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Fionn

Plant cell

Discovery of a Unique Clp Component, ClpF, in Chloroplasts: A Proposed Binary ClpF-ClpS1 Adaptor Complex Functions in Substrate Recognition and Delivery

Kenji Nishimura,a,1 Janina Apitz,c Giulia Friso,a Jitae Kim,a Lalit Ponnala,bBernhard Grimm,c and Klaas J. van Wijka,2

Clp proteases are found in prokaryotes, mitochondria, and plastids where they play crucial roles in maintaining protein

homeostasis (proteostasis). The plant plastid Clp machinery comprises a heterooligomeric ClpPRT proteolytic core, ATPdependent

chaperones ClpC and ClpD, and an adaptor protein, ClpS1. ClpS1 selects substrates to the ClpPR protease-ClpC

chaperone complex for degradation, but the underlying substrate recognition and delivery mechanisms are currently unclear.

Here, we characterize a ClpS1-interacting protein in Arabidopsis thaliana, ClpF, which can interact with the Clp substrate

glutamyl-tRNA reductase. ClpF and ClpS1 mutually stimulate their association with ClpC. ClpF, which is only found in photosynthetic eukaryotes, contains bacterial uvrB/C and YccV protein domains and a unique N-terminal domain. We propose a testable model in which ClpS1 and ClpF form a binary adaptor for selective substrate recognition and delivery to ClpC, reflecting an evolutionary adaptation of the Clp system to the plastid proteome.

Nature biotech

Syngenta rebuffs Monsanto, again

Chinese government reaffirms backing for GM products

Nature

Alternative CRISPR system could improve genome editing

Smaller enzyme may make process simpler and more exact.

Heidi Ledford

Structure of mammalian eIF3 in the context of the 43S preinitiation complex Amedee des Georges, Vidya Dhote, Lauriane Kuhn, Christopher U. T. Hellen, Tatyana V. Pestova+ et al.

The cryo-electron microscopy structure of the eukaryotic initiation factor 3 (eIF3) within the larger 43S complex is determined; the improved resolution enables visualization of the secondary structures of the subunits, as well as the contacts between eIF3 and both eIF2 and DHX29.

Minsoo

1. Proc Natl Acad Sci U S A. 2015 Oct 6;112(40):12456-61. doi: 10.1073/pnas.1517448112. Epub 2015 Sep 21.

Dialogue between E. coli free radical pathways and the mitochondria of C. elegans.

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The microbial world presents a complex palette of opportunities and dangers to animals, which have developed surveillance and response strategies to hints of microbial intent. We show here that the mitochondrial homeostatic response pathway of the nematode Caenorhabditis elegans responds to Escherichia coli mutations that activate free radical detoxification pathways. Activation of C. elegans mitochondrial responses could be suppressed by additional mutations in E. coli, suggesting that C, elegans responds to products of E, coli to anticipate challenges to its mitochondrion. Out of 50 C. elegans gene inactivations known to mediate mitochondrial defense, we found that 7 genes were required for C. elegans response to a free radical producing E. coli mutant, including the **bZip** transcription factor atfs-1 (activating transcription factor associated with stress). An atfs-1 loss-of-function mutant was partially resistant to the effects of free radical-producing E. coli mutant, but a constitutively active atfs-1 mutant growing on wild-type E. coli inappropriately activated the pattern of mitochondrial responses normally induced by an E. coli free radical pathway mutant. Carbonylated proteins from free radical-producing E. coli mutant may directly activate the ATFS-1/bZIP transcription factor to induce mitochondrial stress response: feeding C. elegans with H2O2-treated E. coli induces the mitochondrial unfolded protein response, and inhibition of a gut peptide transporter partially suppressed C. elegans response to free radical damaged E. coli.

2. Nat Cell Biol. 2015 Jun;17(6):782-92. doi: 10.1038/ncb3170. Epub 2015 May 11. A nuclear role for the respiratory enzyme CLK-1 in regulating mitochondrial stress responses and longevity.

Monaghan RM, Barnes RG, Fisher K, Andreou T, Rooney N, Poulin GB, Whitmarsh AJ. Abstract

The coordinated regulation of mitochondrial and nuclear activities is essential for cellular respiration and its disruption leads to mitochondrial dysfunction, a hallmark of ageing. Mitochondria communicate with nuclei through retrograde signalling pathways that modulate nuclear gene expression to maintain mitochondrial homeostasis. The monooxygenase CLK-1 (human homologue COQ7) was previously reported to be mitochondrial, with a role in respiration and longevity. We have uncovered a distinct nuclear form of CLK-1 that independently regulates lifespan. Nuclear CLK-1 mediates a retrograde signalling pathway that is conserved from Caenorhabditis elegans to humans and is responsive to mitochondrial reactive oxygen species, thus acting as a barometer of oxidative metabolism. We show that, through modulation of gene expression, the

pathway regulates both mitochondrial reactive oxygen species metabolism and the mitochondrial unfolded protein response. Our results demonstrate that **a respiratory enzyme acts in the nucleus to control mitochondrial stress responses and longevity.**

3. Proc Natl Acad Sci U S A. 2015 Sep 29;112(39):12093-8. doi: 10.1073/pnas.1515623112. Epub 2015 Sep 11.

Proteomic mapping in live Drosophila tissues using an engineered ascorbate peroxidase.

Chen CL(1), Hu Y(2), Udeshi ND(3), Lau TY(3), Wirtz-Peitz F(2), He L(2), Ting AY(4), Carr SA(3), Perrimon N(5).

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of Genetics, Harvard Medical School, Boston, MA 02115; (3)Broad Institute, Cambridge, MA 02142; (4)Broad Institute, Cambridge, MA 02142; Department of Chemistry, Massachusetts Institute of Technology, Cambridge, MA 02139; (5)Department of Genetics, Harvard Medical School, Boston, MA 02115; Howard Hughes Medical Institute, Boston, MA 02115 clchen@genetics.med.harvard.edu perrimon@receptor.med.harvard.edu.

Characterization of the proteome of organelles and subcellular domains is essential for understanding cellular organization and identifying protein complexes as well as networks of protein interactions. **We established a proteomic**

mapping platform in live Drosophila tissues using an engineered ascorbate peroxidase (APEX). Upon activation, the APEX enzyme catalyzes the biotinylation

of neighboring endogenous proteins that can then be isolated and identified by

mass spectrometry. We demonstrate that APEX labeling functions effectively in multiple fly tissues for different subcellular compartments and maps the mitochondrial matrix proteome of Drosophila muscle to demonstrate the power of APEX for characterizing subcellular proteomes in live cells. Further, we generate "MitoMax," a database that provides an inventory of Drosophila mitochondrial proteins with subcompartmental annotation. Altogether, **APEX labeling in live Drosophila tissues provides an opportunity to characterize the organelle proteome**

of specific cell types in different physiological conditions.

4. Sci Rep. 2015 Sep 2;5:13492. doi: 10.1038/srep13492.

PEA-CLARITY: 3D molecular imaging of whole plant organs.

Palmer WM(1), Martin AP(1), Flynn JR(2), Reed SL(1), White RG(3), Furbank RT(4), Grof CP(1).

Author information:

(1)School of Environmental and Life Sciences, University of Newcastle, Callaghan, NSW, 2308, Australia. (2)School of Biomedical Sciences and Pharmacy, University of Newcastle, Callaghan, NSW 2308, Australia. (3)CSIRO Agriculture, Black Mountain, ACT, 2601, Australia. (4)ARC Centre of Excellence for Translational Photosynthesis, Australian National University, Acton, ACT, 2601, Australia.

Here we report the adaptation of the CLARITY technique to plant tissues with addition of enzymatic degradation to improve optical clearing and facilitate antibody probe penetration. Plant-Enzyme-Assisted (PEA)-CLARITY, has allowed deep

optical visualisation of stains, expressed fluorescent proteins and IgG-antibodies in Tobacco and Arabidopsis leaves. Enzyme treatment enabled penetration of antibodies into whole tissues without the need for any sectioning of the material, thus facilitating protein localisation of intact tissue in 3D whilst retaining cellular structure.

Mary

The network of molecular chaperones: insights in the cellular proteostasis machinery

Marina Ostankovitch^{1,}, Johannes Buchner^{2,}

Keith

Active-State Structures of a Small Heat-Shock Protein Revealed a Molecular Switch for Chaperone Function

Structure 23, 1–10 November 3, 2015

Liu L¹, Chen JY², Yang B³, Wang FH⁴, Wang YH⁵, Yun CH⁶.

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Small heat-shock proteins (sHsps) maintain cellular homeostasis by binding to denatured client proteins to prevent aggregation. Numerous studies indicate that the N-terminal domain (NTD) of sHsps is responsible for binding to client proteins, but the binding mechanism and chaperone activity regulation remain elusive. Here, we report the crystal structures of the wild-type and mutants of an sHsp from Sulfolobus solfataricus representing the inactive and active state of this protein, respectively. All three structures reveal well-defined NTD, but their conformations are remarkably different. The mutant NTDs shows disrupted helices presenting a reformed hydrophobic surface compatible with recognizing client proteins. Our functional data show that mutating key hydrophobic residues in this region drastically altered the chaperone activity of this sHsp. These data suggest a new model in which a molecular switch located in NTD facilitates conformational changes for client protein binding.

Elizabeth

Cell: Alert 3 October-9 October

Andrew P. Bailey, Grielof Koster, Christelle Guillermier, Elizabeth M.A. Hirst, James I. MacRae, Claude P. Lechene, Anthony D. Postle, Alex P. Gould

<u>Antioxidant Role for Lipid Droplets in a Stem Cell Niche of Drosophila</u> Cell, Volume 163, Issue 2, 8 October 2015, Pages 340-353 <u>PDF (0 K)</u> <u>Supplementary</u> <u>content</u>

Luke E. Berchowitz, Greg Kabachinski, Margaret R. Walker, Thomas M. Carlile, Wendy V. Gilbert, Thomas U. Schwartz, Angelika Amon <u>Regulated Formation of an Amyloid-like Translational Repressor Governs</u> <u>Gametogenesis</u>

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Berchowitz et al. establish that transient amyloid-like forms of Rim4, a yeast RNAbinding protein with a predicted prion domain, translationally repress cyclin *CLB3* in meiosis I, thereby ensuring homologous chromosome segregation. These findings suggest that prion domains might enable formation of tightly regulated amyloid-like effectors in diverse functional settings.

A Single α Helix Drives Extensive Remodeling of the Proteasome Lid and Completion of Regulatory Particle Assembly Original Research Article *Pages 432-444* Robert J. Tomko, David W. Taylor, Zhuo A. Chen, Hong-Wei Wang, Juri Rappsilber, Mark Hochstrasser

Highlights

First in vitro reconstitution of RP assembly with completely recombinant components

Electron microscopy and cross-linking reveal massive remodeling of a lid precursor Remodeling of the lid relieves steric clash with the RP base to promote RP assembly Lid remodeling can be triggered by a single C-terminal α helix in the Rpn12 subunit The Plant Journal Content Alert: 84, 2 (October 2015)

Empty pericarp7 encodes a mitochondrial E-subgroup pentatricopeptide repeat protein that is required for *ccmF_N* editing, mitochondrial function and seed development in maize (pages 283–295)

Feng Sun, Xiaomin Wang, Géraldine Bonnard, Yun Shen, Zhihui Xiu, Xiaojie Li, Dahai Gao, Zhonghang Zhang and Bao-Cai Tan

Article first published online: 8 OCT 2015 | DOI: 10.1111/tpj.12993 Significance Statement

Pentatricopeptide repeat (PPR) proteins are important for RNA editing in organelles. Here we show that Emp7 is essential for cytochrome c maturation in mitochondria and consequently for seed development.

<u>Analysis of non-coding transcriptome in rice and maize uncovers roles of</u> <u>conserved lncRNAs associated with agriculture traits (pages 404–416)</u>

Huan Wang, Qi-Wen Niu, Hui-Wen Wu, Jun Liu, Jian Ye, Niu Yu and Nam-Hai Chua Article first published online: 8 OCT 2015 | DOI: 10.1111/tpj.13018 Significance Statement

Long noncoding RNAs (lncRNAs) play key roles in numerous biological processes. Here we sequenced and characterized lncRNAs from many rice and maize tissues.Hundreds of the lncRNAs have single nucleotide polymorphisms associated with agricultural traits, suggesting that they may play roles in controlling such traits

Non-invasive, whole-plant imaging of chloroplast movement and chlorophyll fluorescence reveals photosynthetic phenotypes independent of chloroplast photorelocation defects in chloroplast division mutants (pages 428–442)

Siddhartha Dutta, Jeffrey A. Cruz, Yuhua Jiao, Jin Chen, David M. Kramer and Katherine W. Osteryoung

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Here we demonstrate a dual-imaging platform for continuous, high-sensitivity, and non-invasive measurements of chloroplast movement and chlorophyll fluorescence

in whole plants. We further introduce an approach to correct calculations of nonphotochemical quenching for interference from chloroplast movement. We use this platform and Arabidopsis mutants with drastically enlarged chloroplasts to show that their photosynthetic phenotypes, induced by high-light stress, are due predominantly to altered chloroplast size and shape rather than to reduced chloroplast movement. Plant, Cell & Environment Content Alert (New Articles)

<u>The cytochrome *c* oxidase biogenesis factor AtCOX17 modulates stress responses in</u> <u>Arabidopsis</u>

Lucila Garcia, Elina Welchen, Uta Gey, Agustín L. Arce, Iris Steinebrunner and Daniel H. Gonzalez

Accepted manuscript online: 5 OCT 2015 12:17PM EST | DOI: 10.1111/pce.12647 Current Biology: Alert 30 September-6 October

<u>The Evolutionary Origin of a Terrestrial Flora</u> Review Article *Pages R899-R910* Charles Francis Delwiche, Endymion Dante Cooper

<u>Regulation of Chloroplast Protein Import by the Ubiquitin E3 Ligase SP1 Is</u> <u>Important for Stress Tolerance in Plants</u>

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An Endophyte Constructs Fungicide-Containing Extracellular Barriers for Its Host Plant

Pages 2570-2576 Sameh S.M. Soliman, John S. Greenwood, Aureliano Bombarely, Lukas A. Mueller, Rong Tsao, Dick D. Mosser, Manish N. Raizada

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